

In Silico Analysis of a Chimeric Protein as Alternative Antimicrobial Against Zoonotic Pathogenic Bacteria

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Abstract

The focus of the present study was to characterize chimeric synthetic plantaricin F which naturally produced by *Lactobacillus plantarum* against zoonotic pathogenic bacteria *Staphylococcus aureus* and *Escherichia coli* as antibacterial peptide. The synthetic bacteriocin by bioinformatics revealed higher stability under studied parameter, hence was taken up for further investigation. The amino acids of bacteriocin from *L. plantarum* were analyzed by SnapGene. Further, synthetic PLNF was characterized in silico. The translated partial amino acid sequence of the synthetic PLNF gene displayed 253 amino acids for whole and 148 without tag. The predicted properties of the peptide included theoretical isoelectric point (pI) and hydrophobicity was highly acidic. Molecular weight was 27.2KDa for whole protein and 15.8 KDa for without tag. Prediction of the molecular approach of using SnapGene software and the protein was having antigenicity against bacteria and has B-cell epitope on the surface of protein. Prediction data based on characterization of bacteriocin is novel and predicts synthetic PLNF to be a peptide responsible for antimicrobial activity. The study provides information about a broad spectrum bacteriocin in native probiotic culture and paves the way towards its application as alternative natural antimicrobial agent against zoonotic pathogenic bacteria. Finally, the 3D peptide structure analysis in present study showed that the predicted structure of model and has more functional properties and probably the form most suitable for binding to bacterial cell walls.

Keywords: *Lactobacillus plantarum*, plantaricin, antimicrobial agents, bioinformatics

Introduction

The constant development of bacterial resistance to antibiotic is widely recognized as potential threat to public health. The antimicrobial peptides are broad spectrum and small molecular weight compounds with antagonistic activity against viruses, bacteria and fungi (1). Bacteriocins are antimicrobial peptides presently and beneath consideration equally actual alternatives or complements to community antibiotics (2). Bacteriocin of *Lactobacillus* bacteria are possessive antibacterial activity against species closely related to the producer strains or a wider range of microorganisms (3). The bacteriocin peptides that are produced by *Lactobacillus plantarum* is recognized as plantaricin and is commonly reported as a class II bacteriocin, a very wide-ranging class with a variety of bactericidal and or bacteriostatic mechanisms (4). Plantaricins (A, E, F, J and K) produced by *L. plantarum* (5). Plantaricin F (pln F) is mostly used to prevent microbial contamination (6). Artificial construction gene is the process of synthesizing a gene *in vitro* without the requirement for initial template DNA samples, rapidly step with cost in effect alternative to cloning traditional technique and other molecular biology procedures (7). Moreover, the use of synthetic genes may constitute a successful approach for the heterologous production and functional expression of bacteriocins by recombinant

technique (8). The detection of efficient, safe, and stable agents able to control pathogenic-microorganisms is urgent (9). Bioinformatics is the use of computers in the field of biology to analyze amounts of data and generate new hypothesis. Protein sequence, structure, function and interactions can be studied with some current bioinformatics programs (10). The aim of the present study was bioinformatics analysis of chimeric recombinant plantaricin F that was designed and produced by amino acid optimization approach as antibacterial agent against zoonotic pathogenic bacteria.

Materials and Methods

In the present study, different bioinformatics tools and databases were used for molecular modeling of engineered chimeric protein plantaricin F. Homology modeling procedure was performed in four basic sequential steps: Template selection, target template alignment, model construction, and model assessment. ProSA tool was used to (11).

Design and Construction of Engineered Chimeric Protein

We ordered the nucleotide sequences of desired protein synthesized of plantaricin F (PLNF) gene according to the amino acids sequencing:

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MGSSHHHHHHSSGLVPRGSHMASMTG  
GQQMGRGSEFMSDKIIHLTDDSFDTDV  
LKADGAILVDFWAEWCGPCKMIAPILD
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EIADEYQGKLTVAKLNIDQNPQTAPKY
GIRGIPTLLLFKNGEVAATKVGALSKG
QLKEFLDANLAGSGSGHMHSHHHSS
GLVPRGSGMKETAATAKFERQHMDSPD
LGTDDDDKAMADIGSMKKFLVLRDRE
LNSISGGVFHAYSARGVRNNYKSAVGP
ADWIISAVRGIHG) , as described
previously (12) with some fusion tag (His –
tag , T7 Tag and Thyroxin tag) were
retrieved from the shine/China . The
synthetic fragment was designed with a sites
for two restriction enzyme EcoRI and XhoI
(at 5' and 3' , respectively) with fusion tags
for engineering propose and analysed by
SnapGene (5.4.2) software (13).

Physical and Chemical Parameters

The physical and chemical properties
of synthetic plantaricin F, including its
theoretical isoelectric point (pI), estimated
half-life, instability index, aliphatic index
and lower grand average of hydropath city
(GRAVY).They were predicted by using
ExpASy ProtParam online software
<http://web.expasy.org/protparam/> (14).

B-Cell Epitope Predication

Predict linear B-cell epitopes using IEDB
Analysis Resource at <http://tools.iedb.org/bcell/>
.It is possible to predict linear B-cell epitopes
when threshold at 0.5 as mention previously
(15).

Predication Antigenicity

Predict protein antigenicity of protein was
done by sing VaxiJen server
[http://www.ddgpharmfac.net/vaxijen/VaxiJen/
VaxiJen.html](http://www.ddgpharmfac.net/vaxijen/VaxiJen/VaxiJen.html) (16).

Protein Solubility Predication

Predict protein solubility of synthetic
plantaricin F was done by using
[http://service.tartagliolab.com/new_submission/
ccsol](http://service.tartagliolab.com/new_submission/ccsol) (17).

Three Dimensional Structure Analyses

The 3D of 253 different amino acids of
synthetic PLNF model was done by homology
modelling using chimera (1.15rc-software)
(18) .Inundation , the World wide Protein Data
Bank (wwPDB) which have maintained the
largest, public and free archive of
macromolecular structural data
(www.wwpdb.org) beside the RCSB Protein
Data Bank: which is a powerful new tools for
exploring 3D structures of biological
macromolecules (19) .

Results Protein production was
confirmed by using SnapGene for determine
the amino acids sequencing and fusion tags
(Figure 1).

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AGC AGC GGC CTG GTG CCG CGC GGC AGC CAT ATG GCT AGC ATG ACT
S S G L V P R G S H M A S M T
GGT GGA CAG CAA ATG GGT CGC GGA TCC GAA TTC ATG TCT GAC AAA
G G Q Q M G R G S E F M S D K
ATC ATC CAC CTG ACC GAC GAC TCT TTC GAC ACC GAC GTT CTG AAA
I L H L T D R S F D T D V L A K
GCT GAC GGT GCT ATC CTG GTT GAC TTC TGG GCT GAA TGG TGC GGT
A D G A I L V D F W A E W C G
CCG TGC AAA ATG ATC GCT CCG ATC CTG GAC GAA ATC GCT GAC GAA
P C K M I A P I L D E I A D E
TAC CAG GGT AAA CTG ACC GTT GCT AAA CTG AAC ATC GAT CAG AAT
Y Q G K L T V A K L N I D Q N
CCG GGT ACC GCT CCG AAA TAC GGT ATC CGT GGT ATC CCG ACC CTG
P G T A P K Y G I R G I P T L
CTG CTG TTC AAA AAC GGT GAA GTT GCT GCT ACC AAA GTT GGT GCT
L L F K N G E V A & T K V G A
CTG TCT AAA GGT CAG CTG AAA GAA TTT CTG GAC GCT AAC CTG GCT
L S K G Q L K E F L D A N L A
GGC TCC GGC TCT GGT CAC ATG CAC CAT CAC CAT CAC CAT TCT TCT
G S G S G H M H H H H H H S S
GGT CTG GTT CCG CGT GGT TCT GGT ATG AAA GAA ACC GCT GCT GCT
G L V P R G S G M K E T A & &
AAA TTC GAA CGT CAG CAC ATG GAC TCT CCG GAC CTG GGT ACC GAC
K F E R Q H M D S P D L G T D
GAC GAT GAT AAA GCT ATG GCT GAC ATC GGT TCT ATG AAA AAA TTC
D D D K A M A D I G S M K & & F
CTG GTT CTG CGT GAC CGT GAA CTG AAC TCT ATC TCT GGT GGT GTT
L V L R D R E L N S I S G G G L G V
TTC CAC GCT TAC TCT GCT CGT GGT GTT CGT AAC AAC TAC AAA TCT
F H A V S A R G V R N X V K S
GCT GTT GGT CCG GCT GAC TGG ATC ATC TCT GCT GTT CGT GGT TTC
A V G P A D W I L S A V R G F
ATC CAC GGT TAG CTGAG
I H G *

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Figure1: Nucleotide and amino Acid sequence of fused Recombinant synthetic PLNF analysis by SnapGene, The Gray color is thioredoxin-(His)6tag fusion partner protein followed with S-Tag and enterokinase cleavage site (1-165 aa), the Red color is leader peptide of pln F (166-183 aa), the Brown color is mature peptide of pln F (184-217 aa), the asterisk is stop codon.

● His tag ● Thrombin site ● T7 tag ● EcoRI ● Trx tag ● S tag ● Enterokinase site ● The leader peptide of pln F ● The mature peptide of plnF ● Start / Stop codon, respectively ● XhoI

Physical and Chemical Parameters

Synthetic plantaricin was predicted to be a stable, molecular 27.2 KDa according to SnapGene, chimeric protein with an estimated half-life of more than 10 h in *E. coli*. The aliphatic and grand averages of hydropathicity (GRAVY) indices were to be for whole bacteriocin - 0.364 and -0.228- respectively- The physicochemical properties of the protein and the results showed in table 1.

B-Cell Epitope Prediction for Synthetic Plantaricin F

The protein sequence was analyzed using Bepipred linear epitope prediction, surface accessibility and to assess the antigenicity respectively. The average for the whole protein was 0.520 and without tag was 0.489. All values equal or greater than the default threshold 0.5 were predicted to be a potential B cell binder (Figure 2 and 3). The

yellow region was a positive prediction of B cell epitope and the green region was negative.

PLNF protein. The threshold for these proteins was considered 0.4 and the synthetic was more than 0.4 (Table2).

Predict The Antigenicity

VaxiJen v2.0 was used as bioinformatics - tool to analyze antigenicity index of synthetic

Table 1: The analysis result physicochemical properties of synthetic PLNF protein.

Properties	Whole construct	Without tags
Number of amino acids	253	148
Molecular weight	27285.80	15878.06
Theoretical pI	6.64	6.11
Formula	C ₁₁₉₂ H ₁₈₆₁ N ₃₅₁ O ₃₆₀ S ₁	C ₆₉₉ H ₁₀₉₆ N ₁₉₆ O ₂₁
Total number of atoms	3777	2210
<i>in vitro</i>	30 hours	30 hours
Estimated Yeast	>20 hours	>20 hours
half-life <i>Escherichia coli</i>	>10 hours	>10 hours
Instability index (state)	19.10 (stable)	17.53 (stable)
Aliphatic index	73.72	81.15
Grand average of hydropathcity GRAVY	-0.364	-0.228

Table 2: Antigenicity index synthetic PLNF protein

Properties	Whole construct	Without tags
Overall Prediction for the protective antigen (antigen/non-antigen)	0.4867 (Probable ANTIGEN)	0.5303 (Probable ANTIGEN)

Predict The Solubility

Predict the solubility propensity of the purified the ccSol was applied. The possible expect results were demonstrated -solubility of synthetic PLNF was 84% with tag and 96% without tag (Figures 4 and 5). Moreover, the ccsol omics bioinformatics website was given the probable results to predict the solubility of expression PLNF was 92% with tag and 98% without tag (Figures 6 and 7).

Three Dimensional Structures

The amino acid sequences of 253 chimeric synthetic plantaricin F was obtained using an

in silico sequence conversion tool. The amino acid sequences were analyzed by SnapGene software. The 3D structures of 253 chimeric proteins were compared with the 3D structures using Chimera (Figure 8). The 3D qualified structure prediction of the chimeric protein was detected using the Robetta server tool <https://robetta.bakerlab.org/> (Figure 9). Figure 10 and 11 were provided qualified 3D structures of the synthetic PLNF protein. The UCSC Chimera was used to measure the root-mean-square deviation of atomic positions (RMSD) between the whole protein and the recombinant synthetic PLNF protein without the mentioned tags (Figure 12).

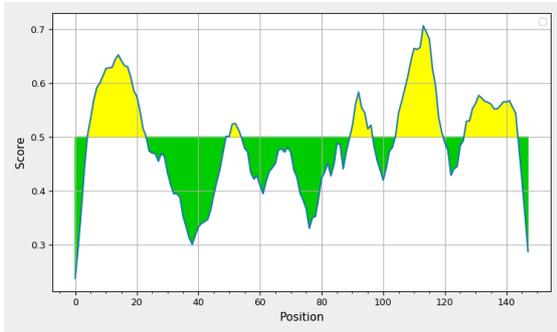


Figure 3: The linear B-cell epitopes results for protein without tags. Average: 0.489 , Minimum: 0.238 Maximum: 0.706

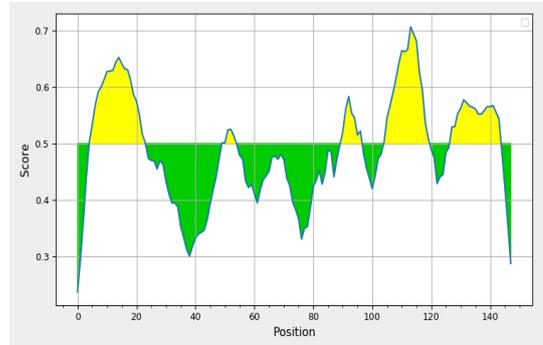


Figure 2: The linear B-cell Epitopes results for the whole protein. Average: 0.520 Minimum: 0.253 Maximum: 0.713.

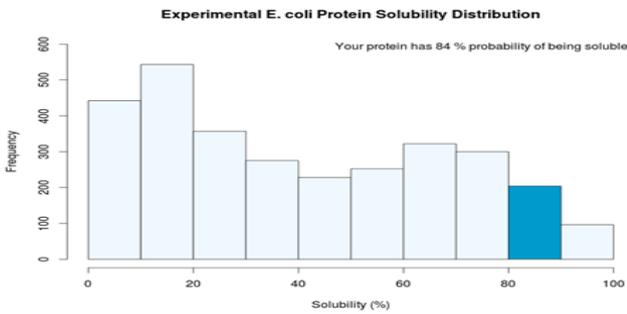


Figure5 : Solubility propensity results for the whole Synthetic PLNF

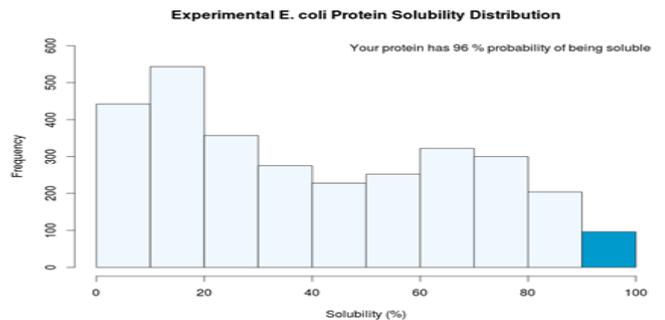


Figure 4: Solubility propensity for synthetic PLNF without the tags.

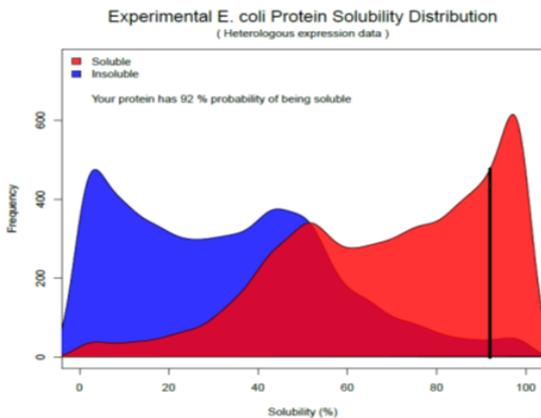


Figure7: The probable solubility for synthetic PLNF without tags. Inset: Overall score distribution for soluble (red) and insoluble (blue) proteins

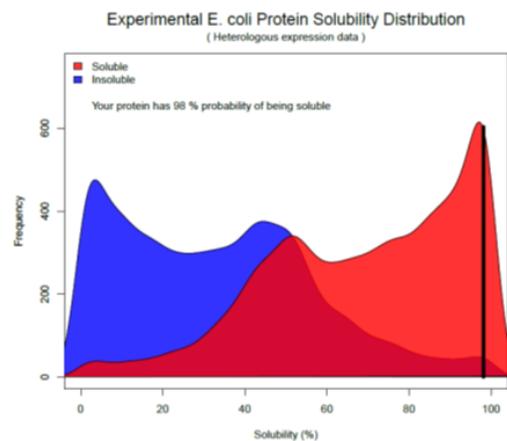


Figure 6: The probable solubility for synthetic PLNF with Tags. Inset: Overall score distribution for soluble (red) and insoluble (blue) proteins



Figure9: Protein 3D structure alignments generated by the UCSF Chimera package: The 3D structure in cartoon mode whole synthetic PLNF as modeled by Robetta server. The cartoon structure color was cyan.

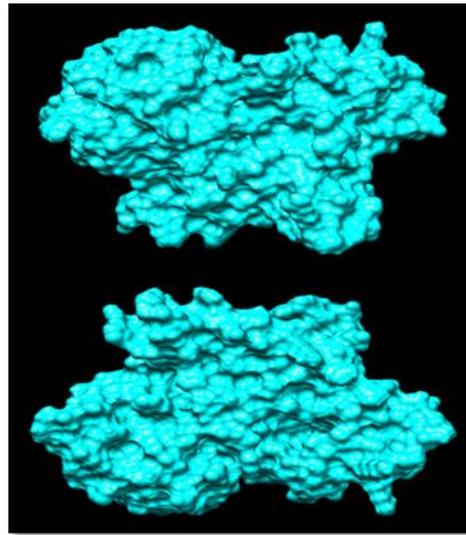


Figure 8: The 3D structure of surface mode for whole synthetic PLNF as modeled by Robetta server. The structure was staining in cyan color .

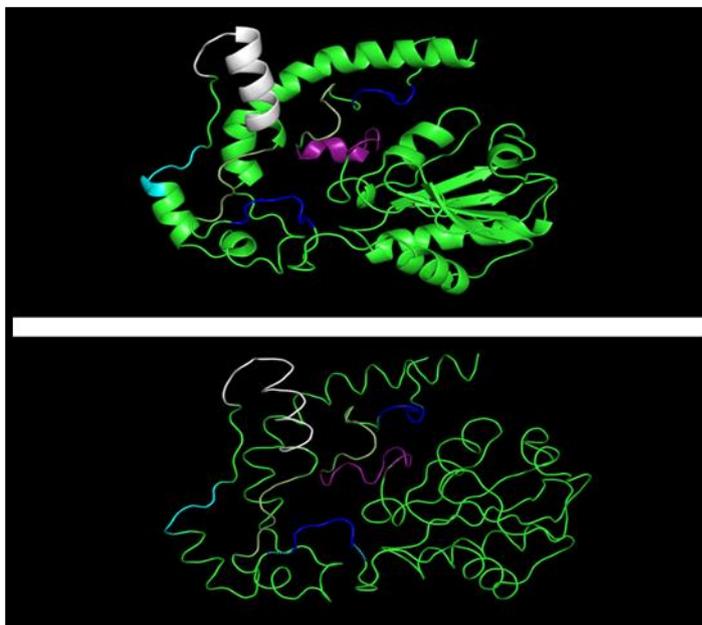


Figure10. The top structure in cartoon mode and the bottom structure is Ribbon mode explains the 3D structure of the synthetic PLNF as modeled by Robetta server. His-tag in red, thrombin site in smudge, T7 tag in deep purple, S-tag in white, enterokinase site in cyan, and the rest of the protein in green color.

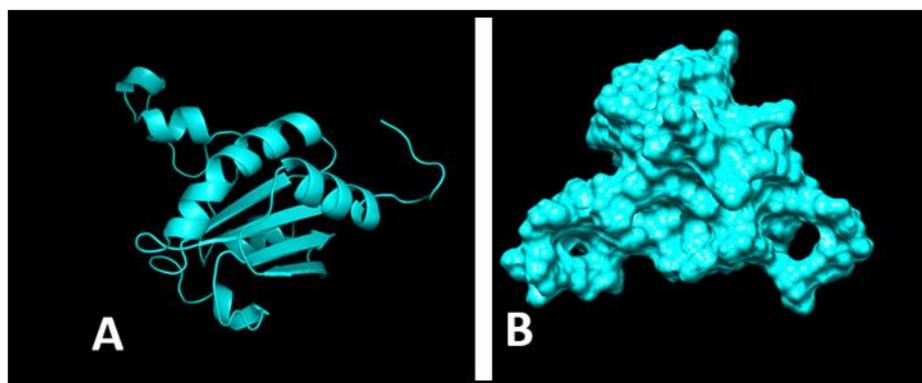


Figure 11: The 3D structure of the synthetic PLNF without the tags as modeled by Robetta server. The (A) structure was in cartoon mode However, the (B) structure represented the surface mode. The color is cyan

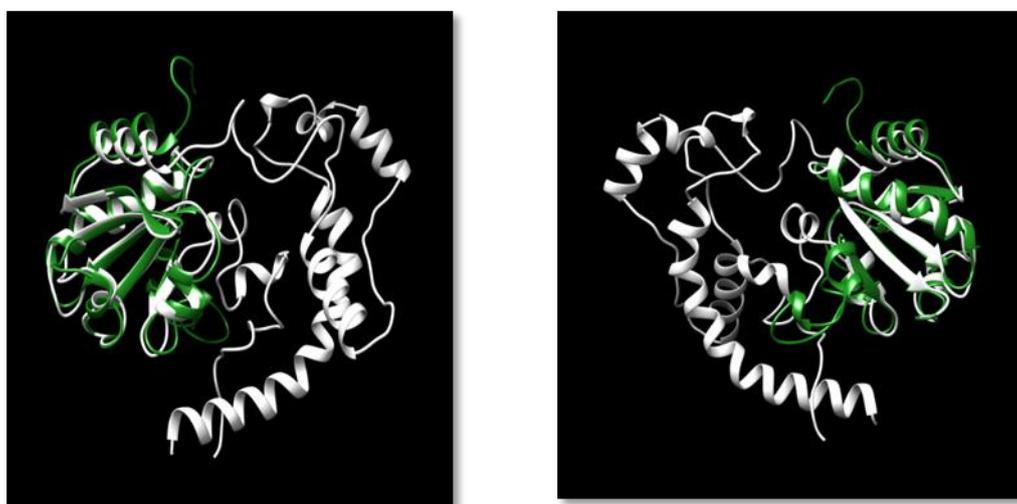


Figure 12: The figures were the same but in different angles. The measured RMSD1 was 1.12 angstrom which was indicative of no substantial changes in the 3D Structure of the whole protein while harbouring the Tags. The Structures are in Ribbon mode, and the whole protein represented in white while the protein without the tags is in green. In bioinformatics, the root-mean-square deviation of atomic positions, or simply root-mean-square deviation (RMSD), is the measure of the average distance between the atoms (usually the backbone atoms) of superimposed proteins.

Discussion:

Recombinant DNA technical methods have permitted fusions of genes in a simple way (20). Designing recombinant protein via bioinformatics approach is comparatively stable, safe, inexpensive, specific and more effective. Thus, herein, we made a chimeric protein plantaricin F for the 1st time. The designed fusion synthetic plantaricin *F* gene was constructed and cloned into pUC57 vector and transformed into cloning host cell *E. coli* BL21(21).

The results of protein analysis of sequence of amino acid of synthetic PLNF were investigated by using the SnapGene software, SnapGene allows one to view the default sequence, commercially tag sequence classes, and change the view settings (Figure 1). The allowance of such capabilities might be integrated with much larger data sets or genotypes and parallel processing may help offload latency times for creating output visualization (22). The mature sequence VFHAYSARGVRNNYKSAVGPADWIISAV RGFHIG of synthetic PLNF was similarity 100% to pervious study of (23). However, fusion tags include His tag, T7 tag, Trx tag and S tag used in *E. coli* to improve protein production yields, solubility and folding, and to facilitate protein purification. They can also confer specific properties for target proteins characterization. Fusion partners can also be used when producing toxic proteins for

production of antimicrobial peptides by *E. coli* agree with Costa *et al.*, (24). On the other hand, the ProtParam analysis showed that detected the synthetic PLNF sequence, in which the molecular weight was calculated by the addition of average isotopic masses of amino acids, the molecular weight was 27.2 KDa which provides information on the full-length of protein expression, expression of modified, and cleavage products (Table 1). The instability index showed stable in nature considered to be stable (instability index <40). The theoretical isoelectric point (pI) was less than 7 (pI < 7) which indicates synthetic PLNF was considered highly acidic in nature. The computed isoelectric point will be useful for developing a buffer system for purification by isoelectric focusing method (25). The hydrophilic nature of the protein and the possibility of better interaction with water indicated by GRAVY indices value (-0.364, -0.288) was highly acidic (Table 2) (26).

The results antigenicity revealed all values were more than the threshold 0.4 (Table 3). This finding agreement with previously study (27). It has been found that synthetic PLNF have antigenicity against bacteria. While, the synthetic PLNF showed a high solubility (Figures 3 and 4) which indicate another positive feature of the synthetic PLNF that prevents the accumulation of protein in the inclusion and was used to confirm the full-

length of synthetic PLNF suitability for expression in *E. coli* and solubility propensity also indicated that a less accumulation of proteins in the inclusion bodies with the addition of restriction enzymes and a N-terminal his tag, as shown in figures 5 and 6, which also revealed the probable solubility was high heterologous expression for synthetic PLNF (red) soluble in compare with insoluble (blue) protein, and this finding agree with (20,28). On other hand, the graphic revealed all values equal to or greater than the threshold at 0.5. In which the Y-axis defines for each amino acid residual score, however, X is the position of the amino acid sequence residue (Figures 7 and 8) The results were showed specific areas in proteins that bind to the B cell receptor (29). Additionally, UCSF chimera was an extensible program for interactive visualization and analysis of molecular structures. The 3D peptide structure analysis in present study showed that the predicted structure of model has more functional properties and probably the form most suitable for binding to bacterial cell walls (30).

Conclusion

Bioinformatics approaches have ability to analyze bacteriocin avoiding the disadvantages of the conventional methods reduce time, cost required, enhance the safety and efficacy of the predicted epitopes. Several epitopes were predicted in the current study to study chimeric

synthetic PLNF against zoonotic pathogenic bacteria using in silico prediction tools. Bioinformatics is recommended for further *in vitro* studies for confirmation and synthetic PLNF can be used as alternative antibacterial agents against pathogenic bacteria in medicine and veterinary medicine.

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Conflict of Interest

The authors state that there is no conflict of interest.

References

1. Yang, L. M.; Shu, C.; Wu, J.; Zhao, N. and Lu, Z. L. (2013). Circular function-based gas-kinetic scheme for simulation of inviscid compressible flows. *Journal of Computational Physics*, 255, 540–557.
2. Chikindas, M. L.; Weeks, R., Drider, D.; Chistyakov, V. A.; and Dicks, L. M. (2018). Functions and emerging applications of bacteriocins. *Current opinion in biotechnology*, 49, 23-28.
3. Wang, B.; Lucy, K. A.; Schuman, J. S.; Sigal, I. A.; Bilonick, R. A.; Lu, C., Liu, J.; Grulkowski, I.; Nadler, Z. and Ishikawa, H. (2018). Tortuous pore path through the

- glaucomatous lamina Cribrosa. *Scientific Reports*, 8(1), 1–7.
4. Ahmad, V.; Khan, M. S.; Jamal, Q. M. S.; Alzohairy, M. A.; Al Karaawi, M. A.; and Siddiqui, M. U. (2017). Antimicrobial potential of bacteriocins: in therapy, agriculture and food preservation. *International Journal of Antimicrobial Agents*, 49(1), 1–11.
 5. Kjos, M.; Nes, I. F. and Diep, D. B. (2009). Class II one-peptide bacteriocins target a phylogenetically defined subgroup of mannose phosphotransferase systems on sensitive cells. *Microbiology*, 155(9), 2949–2961.
 6. Mustopa, A. Z.; Murtiyaningsih, H.; Fatimah, F. and Suharsono, S. (2016). Cloning and heterologous expression of extracellular plantaricin F produced by *Lactobacillus plantarum* S34 isolated from “Bekasam” in *Lactococcus lactis*. *Microbiology Indonesia*, 10(3), 3
 7. Schwartz, M.; Lo, H.; Zhou, J.; Yang, P. and Liao, A. (2011). Gene Synthesis: A Cost-Effective Alternative to Traditional Molecular Cloning. *Journal of Biomolecular Techniques: JBT*, 22(Suppl), S31.
 8. Jiménez, J. J.; Borrero, J.; Gútierez, L.; Arbulu, S.; Herranz, C.; Cintas, L. M. and Hernández, P. E. (2014). Use of synthetic genes for cloning, production and functional expression of the bacteriocins enterocin A and bacteriocin E 50-52 by *Pichia pastoris* and *Kluyveromyces lactis*. *Molecular Biotechnology*, 56(6), 571–583.
 9. Radošević, K.; Čanak, I.; Panić, M.; Markov, K.; Bubalo, M. C.; Frece, J.; Srček, V. G. and Redovniković, I. R. (2018). Antimicrobial, cytotoxic and antioxidative evaluation of natural deep eutectic solvents. *Environmental Science and Pollution Research*, 25(14), 14188–14196.
 10. Amela, I.; Delicado, P.; Gómez, A.; Querol, E.; and Cedano, J. (2013). A dynamic model of the proteins that form the initial iron-sulfur cluster biogenesis machinery in yeast mitochondria. *The protein journal*, 32(3), 183-196.
 11. Wiederstein and Sippl (2007) Wiederstein, M. and Sippl, M. J. (2007). ProSA-web: interactive web service for the recognition of errors in three-dimensional structures of proteins. *Nucleic Acids Research*, 35(suppl_2), W407–W410.
 12. Kusdianawati, K.; Mustopa, A. Z.; Suharsono, S.; Budiarto, B. R.; Fatimah, F. and Danuri, H. (2015). Construction, expression and purification of recombinant pre-mature peptide of plantaricin F from *Lactobacillus plantarum* S34 in *Escherichia coli*. *Indonesian Journal of Agricultural Science*, 16(1), 31-38.
 13. Khalil, M. I. (2020, November). CRISPRCas9 design to knockout and

- knockin the breast cancer gene-BRCA1 in *Arabidopsis thaliana*. In *Journal of Physics Conference Series*, 1660, (1), 012009. IOP Publishing.
14. Gasteiger, E.; Gattiker, A.; Hoogland, C.; Ivanyi, I.; Appel, R. D. and Bairoch, A. (2003). ExPASy: the proteomics server for in-depth protein knowledge and analysis. *Nucleic Acids Research*, 31(13), 3784–3788.
 15. Collatz, M.; Mock, F.; Barth, E.; Hölzer, M.; Sachse, K. and Marz, M. (2021). EpiDope: A Deep neural network for linear B-cell epitope prediction. *Bioinformatics*, 37(4), 448–455.
 16. Doytchinova, I. A. and Flower, D. R. (2007). VaxiJen: a server for prediction of protective antigens, tumor antigens and subunit vaccines. *BMC bioinformatics*, 8(1), 1-7.
 17. Agostini, F., Cirillo, D., Livi, C. M., Delli Ponti, R. and Tartaglia, G. G. (2014). cc SOL omics: A webserver for solubility prediction of endogenous and heterologous expression in *Escherichia coli*. *Bioinformatics*, 30(20), 2975-2977.
 18. Pettersen, E. F., Goddard, T. D., Huang, C. C., Couch, G. S., Greenblatt, D. M., Meng, E. C. and Ferrin, T. E. (2004). UCSF Chimera—a visualization system for exploratory research and analysis. *Journal of computational chemistry*, 25(13), 1605-1612.
 19. Burley, S. K., Bhikadiya, C., Bi, C., Bittrich, S., Chen, L., Crichlow, G. V. and Zhuravleva, M. (2021). RCSB Protein Data Bank: powerful new tools for exploring 3D structures of biological macromolecules for basic and applied research and education in fundamental biology, biomedicine, biotechnology, bioengineering and energy sciences. *Nucleic acids research*, 49(D1), D437-D451.
 20. Haghroosta, A., Goudarzi, H., Faghihloo, E., Ghalavand, Z., Ranjbar, M. M. and Langroudi, R. P. (2020). In silico analysis of a chimeric fusion protein as a new vaccine candidate against *Clostridium perfringens* type A and *Clostridium septicum* alpha toxins. *Comparative clinical pathology*, 29(5), 981-989.
 21. Dana, H., Chalbatani, G. M., Gharagouzloo, E., Miri, S. R., Memari, F., Rasoolzadeh, R. and Marmari, V. (2020). In silico analysis, molecular docking, molecular dynamic, cloning, expression and purification of chimeric protein in colorectal cancer treatment. *Drug design, development and therapy*, 14, 309.
 22. Nassief, A. M. (2020). Conventional Data Science Techniques to Bioinformatics and Utilizing a Grid Computing Approach to Computational Medicine.
 23. Widyasari, E. M., Mustopa, A. Z., Daruwati, I., Sriyani, M. E., Sugiharti, R. J., and

-
- Gurning, E (2018). Radiolabing of plantaricin F as a natural antibiotics using iodine -131. *Journal of Pharmaceutical Sciences and Research*, 10(11), 72-87.
24. Costa, S., Almeida, A., Castro, A. and Domingues, L. (2014). Fusion tags for protein solubility, purification and immunogenicity in *Escherichia coli*: the novel Fh8 system. *Frontiers in microbiology*, 5, 63.
25. Sahay, A., Piprodhe, A. and Pise, M. (2020). In silico analysis and homology modeling of strictosidine synthase involved in alkaloid biosynthesis in *catharanthus roseus*. *Journal of Genetic Engineering and Biotechnology*, 18(1), 1-6.
26. Saikat, A. S. M. (2020). Structure Prediction and Characterization of Uncharacterized ABC Transporter ATP-Binding Protein Rv0986 of *Mycobacterium tuberculosis* (Strain ATCC 25618/H37Rv). *bioRxiv*.112680.
27. Ghahremanifard, P., Afzali, F., Rostami, A., Nayeri, Z., Bambai, B. and Minuchehr, Z. (2020). Designing a novel multi-epitope T vaccine for “targeting protein for Xklp-2” (TPX2) in hepatocellular carcinoma based on immunoinformatics approach. *International Journal of Peptide Research and Therapeutics*, 26(2),1127-1136
28. Yazdani, Z., Rafiei, A., Valadan, R., Ashrafi, H., Pasandi, M., and Kardan, M. (2020). Designing a potent L1 protein-based HPV peptide vaccine: a bioinformatics approach. *Computational biology and chemistry*, 85, 107209.
29. Alam, S., Hasan, M. K. and Hossain, M. F. (2018). Analysis of Maize Profilin-4 Isoform as an allergen. *bioRxiv*, 503425.
30. Tanhaeian, A.; Damavandi, M. S.; Mansury, D. and Ghaznini, K. (2019). Expression in eukaryotic cells and purification of synthetic gene encoding enterocin P: a bacteriocin with broad antimicrobial spectrum. *AMB Express*, 9(1), 1-9 .

تحليل السيليكو لبروتين مهجن كمضاد بكتيري بديل ضد البكتيريا المسببة للأمراض في الانسان و الحيوان

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الخلاصة

تركزت الدراسة الحالية على توصيف بلانترسين F الكيميري الاصطناعي الذي ينتج بشكل طبيعي بواسطة *Lactobacillus plantarum* ضد البكتيريا المسببة للأمراض في الانسان والحيوان *Staphylococcus aureus* و: *Escherichia coli* كبيتيد مضاد. أظهرت النتائج من خلال دراسة المعلوماتية الحيوية ثباتاً أعلى للبيتيد مع الواسم المستخدم. تم تحليل الأحماض الأمينية للبكتريوسين من *L. plantarum* بواسطة SnapGene. علاوة على ذلك ، تم تمييز PLNF الاصطناعي في السيليكو. تم اجراء تسلسل للأحماض الأمينية الجزئية المترجمة لجين PLNF الاصطناعي، فكانت 253 حامض اميني للبيتيد الكامل مع الواسم و 148 حامض اميني للبيتيد الكامل بدون واسم . أظهرت النتائج انه بيتيد شديدة الحموضة. كان الوزن الجزيئي للبكتريوسين الكامل 27.2 كيلو دالتون مع الواسم اما بدونه فكان 15.8 كيلودالتون. يتوقع أن يكون PLNF الاصطناعي عبارة عن بيتيد مسؤول عن نشاط مضاد واسع للميكروبات. توفر هذه الدراسة معلومات حول مجموعة واسعة من البكتريوسين التي لها فعالية حيوية ضد الميكروبات وتمهد الطريق نحو استخدام هذا البيتيد كعامل طبيعي بديل لمضاد للميكروبات ضد البكتيريا المسببة للأمراض في الانسان والحيوان . أخيراً ، أظهر التحليل ثلاثي الابعاد للبيتيد في دراستنا أن له خصائص وظيفية متعددة وربما يكون الشكل الأكثر ملائمة للارتباط بجدران الخلايا البكتيرية ومن ثم تدميرها.

الكلمات المفتاحية : بلانترسين، العوامل المضادة للميكروبات ، المعلوماتية

Lactobacillus plantarum